

Effect of calcium on acid secretion by the isolated rat gastric mucosa

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Changes in plasma Ca^{++} levels have been shown to affect gastric acid secretion in man and other species (Barreras, 1973). These effects may be caused indirectly via hormonal mechanisms or by a direct action on the parietal cells. We have therefore investigated the effects of various external Ca^{++} concentrations on secretory responses to histamine and to pentagastrin using the isolated rat gastric mucosa (Hearn & Main, 1975).

Ca^{++} concentrations were adjusted over the range of 0.0 to 7.2 mM, in both mucosal and serosal solutions, keeping other cations constant. Drugs were added serosally and acid output recorded via an electrode in the unbuffered mucosal solution. An initial control response (in 3.6 mM Ca^{++}) was compared with the mean of two responses, 90 min apart, in the presence of altered Ca^{++} .

When the control conditions (3.6 mM Ca^{++}) were maintained throughout the 450 min experimental period, a progressive decrease in basal secretion rate was seen. This trend was increased by 7.2 mM Ca^{++} and reversed by 0.9 or 0.0 mM Ca^{++} . The effects were readily reversed on washing with 3.6 mM solutions.

Ca^{++} produced similar effects on acid responses to histamine (25×10^{-6} M, 45 min contact). In control experiments, the responses increased slightly ($+0.55 \pm 0.46 \mu\text{mol cm}^{-2} \text{h}^{-1}$, mean change \pm s.e. mean), while lowering Ca^{++} to 1.8 or 0.9 mM produced larger changes ($+3.68 \pm 0.41$, $P < 0.002$ and $+4.67 \pm 1.04$, $P < 0.01$ respectively). In the presence of Ca^{++} -free solutions, with EDTA (0.5 mM) in serosal

solution only, test responses were not significantly altered from control ($+0.42 \pm 0.84$, $n=8$ for all groups).

Using the same experimental design, consistent responses to pentagastrin (1.8×10^{-8} M, 30 min contact) were obtained. Control responses to pentagastrin and histamine did not differ significantly (1.18 ± 0.24 , $n=25$ and 2.01 ± 0.47 , $n=32$, respectively). Under control conditions, there was a mean fall in the test responses to pentagastrin of $-0.17 \pm 0.08 \mu\text{mol cm}^{-2} \text{h}^{-1}$ with a larger decrease of -0.61 ± 0.35 being produced by 7.2 mM Ca^{++} ($n=6$). A significant increase in size of responses was observed with both 0.9 mM and Ca^{++} -free solutions ($+0.95 \pm 0.29$, $P < 0.05$, $n=6$ and $+1.04 \pm 0.15$, $P < 0.002$, $n=7$ respectively).

Raising Mg^{++} concentrations from 1.2 to 2.4 and 4.8 mM (keeping Ca^{++} constant at 3.6 mM) caused a potentiation of responses to histamine, the results being significant only for 2.4 mM solutions ($+4.97 \pm 1.79$, $P < 0.05$, $n=5$).

These results demonstrate that, in the rat *in vitro*, lowering external Ca^{++} increases both basal and stimulated acid secretion by a direct effect on the secretory mucosa. They provide little support for the hypothesis that pentagastrin acts via Ca^{++} -dependent histamine release within the mucosa, since changing the Ca^{++} concentration had similar effects on the responses to histamine and pentagastrin.

J.B.P. is an M.R.C. Student.

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The effects of some gastrointestinal hormones and metoclopramide on cardiovascular dopamine receptors

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During investigations of the pharmacology of metoclopramide, a drug known to accelerate gastric

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emptying and stimulate intestinal motility in animals (Jacoby & Brodie, 1967) and man (James & Hume, 1968), we have considered the possibility of an interaction between metoclopramide and various hormones which affect gastrointestinal motility. Metoclopramide blocks cardiovascular dopamine receptors (Day & Blower, 1975) and some actions of secretin may be mimicked by dopamine (Furuta, Hashimoto, Iwatsuki & Takeuchi, 1973). We have therefore investigated the effects of secretin, glucagon, cholecystokinin and the synthetic peptide pentagastrin on cardiovascular dopamine receptors in anaesthetized rats.

Anaesthetized male Wistar rats were prepared as previously described (Day & Blower, 1975). After α - and β -adrenoceptor blockade, the intravenous

administration of dopamine (0.1 mg/kg), secretin (0.2 u/kg), glucagon (0.5 µg/kg) or cholecystokinin (0.15 u/kg) elicited short-lived falls in blood pressure. These depressor responses were similar in magnitude and duration to those of dopamine. Responses to secretin and glucagon were rapidly tachyphylactic, but this effect was reversed following administration of dopamine and could be prevented by alternating doses of dopamine with doses of either hormone. Responses to cholecystokinin did not show tachyphylaxis but did reduce the responses to subsequent doses of dopamine, secretin or glucagon.

Pentagastrin (0.1 µg/kg) evoked short-lived increases in blood pressure. The hypotensive responses to dopamine, secretin, glucagon and cholecystokinin were completely reversed following administration of pentagastrin (0.1 µg/kg) or metoclopramide (5 mg/kg).

These results suggest that secretin and glucagon may have direct or indirect agonist actions at dopamine receptors. Cholecystokinin appeared to behave as a partial agonist, whilst both pentagastrin

and metoclopramide acted as dopamine receptor antagonists. The interaction of these hormones with dopamine receptors could be of importance in elucidating some of their effects on vascular haemodynamics or on gastrointestinal motility.

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The effects of histamine and selective histamine receptor agonists on the isolated working guinea-pig heart preparation

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We have recently described an isolated guinea-pig heart preparation capable of sustaining external work (Flynn, Gristwood & Owen, 1977). Using this preparation the effects of histamine and the selective histamine H_1 - and H_2 -receptor agonists, 2-pyridylethylamine (Durant, Ganellin & Parsons, 1975) and dimaprit (Parsons, Owen, Ganellin & Durant, 1977) respectively, on cardiac function were investigated.

Full dose response curves for each agonist were constructed in separate preparations. Parallel line assays were used to compare the potencies of 2-pyridylethylamine and dimaprit with histamine. The analysis of variance used to estimate potency showed that the dose-response curves to each of the agonists were parallel over the dose-range used.

Control values for each measured parameter after stabilization of the preparations were: external work (E.W.) 0.410 ± 0.017 kg-m min⁻¹ g⁻¹, maximum left ventricular pressure (L.V.P. max.) 84.5 ± 1.2 mmHg,

contractility as indicated by dL.V.P./dt max. (dp/dt max.) 2239 ± 96 mmHg/s, sinus rate (S.R.) 216.2 ± 4.4 beats/min, aortic flow (A.F.) 360.0 ± 13.8 ml min⁻¹ g⁻¹, coronary flow (C.F.) 73.0 ± 4.1 ml min⁻¹ g⁻¹ and cardiac output (C.O.) 433.1 ± 16.8 ml min⁻¹ g⁻¹, ($n = 20$). Where appropriate, measurements are expressed per gram dry weight of heart. These and subsequent figures are means \pm s.e. mean.

Histamine increased all measured parameters of cardiac function over the dose-range 10^{-9} to 10^{-6} mol. The maximum absolute increase in each parameter, was calculated from the equation $V = (Vm/(1 + K)/A$, where V = response, Vm = maximum response, K = dissociation constant of agonist and A = dose of agonist. The increases were: E.W. 0.353 ± 0.026 kg-m min⁻¹ g⁻¹, L.V.P. max. 41.5 ± 2.0 mmHg, dp/dt max. 4132 ± 229 mmHg/s, S.R. 122.9 ± 4.9 beats/min, A.F. 152.7 ± 14.6 ml min⁻¹ g⁻¹, C.F. 42.9 ± 4.1 ml min⁻¹ g⁻¹ and C.O. 188.8 ± 11.9 ml min⁻¹ g⁻¹, ($n = 8$).

Dimaprit produced similar changes to histamine in all parameters over the same dose-range. The maximum increase in each parameter was: E.W. 0.422 ± 0.035 kg-m min⁻¹ g⁻¹, L.V.P. max. 48.4 ± 6.1 mmHg, dp/dt max. 4398 ± 340 mmHg/s, S.R. 122.8 ± 7.8 beats/min, A.F. 174.5 ± 19.6 ml min⁻¹ g⁻¹, C.F. 41.6 ± 1.1 ml min⁻¹ g⁻¹ and C.O. 211.5 ± 20.1 ml min⁻¹ g⁻¹, ($n = 6$).

The potencies, with fiducial limits, of dimaprit relative to histamine (100%) for each parameter, were: E.W. 74.4 (57.2-96.8)%, L.V.P. max. 47.0